

Molecular detection of protozoan parasite infections in marine bivalves in Korean waters

Hye-Mi Lee¹, Young-Ghan Cho¹, Jeong-Hwa Kim¹, Jong-Seop Shin¹, Kajino Nobuhisa¹, Min-Seok Jang², Jee Youn Hwang², and Kwang-Sik Choi¹

¹Department of Marine Life Science (BK21 FOUR), Jeju National University
²Aquatic Disease Control Division, National Institute of Fisheries Science of Korea

Backgrounds and Objective

- Several species of protozoan parasites, including the members in the genus *Perkinsus*, *Marteilia*, and *Bonamia* are listed and regulated by the world organization for animal health (OIE), as they often cause mass mortalities of the host organisms.
- In Korea, small bays on the south coast are used as major culture grounds of the Pacific oysters (*Crassostrea gigas*), where approximately 150,000 MTs of oysters are produced annually.

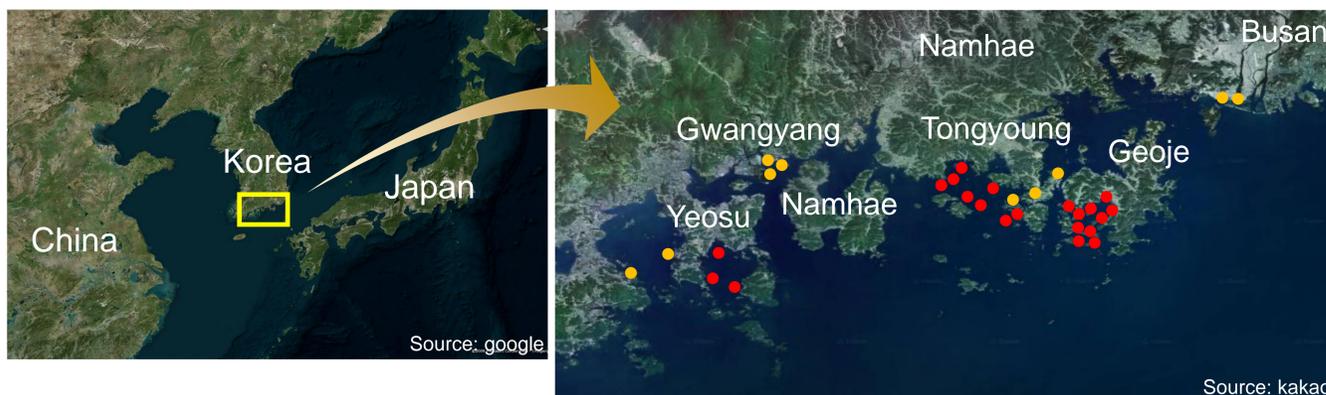


Suspended long-line culture system in oyster grow-out fields and harvesting oysters using an automatic oyster-string processor

- The Pacific oysters on the south coast of Korea are known to be infected by *Marteillioides chungmuensis*, while infection by other OIE-listed protozoan parasites is unknown.
- In this study, possible infections by *Bonamia ostreae*, *B. exitiosa*, *Marteilia refringens*, *Perkinsus marinus*, and *P. olseni* in the Pacific oysters from the south coast of Korea.

Materials and Methods

- For the analysis, commercially raised (N=1,260) and wild (N=600) Pacific oysters were collected from 31 sites on the south coast in spring (pre-spawning season) and fall (post-spawning season) 2020.



- Commercial culture area (21 sites)
- Intertidal area (10 sites)

- For the analysis, commercially raised (N=1,500) and wild (N=360) Pacific oysters were collected from 31 sites on the south coast in spring (pre-spawning season) and fall (post-spawning season) 2020.
- The OIE-listed five protozoan parasites in the oysters was examined using PCR with the species-specific primers recommended by the OIE.

Species	Primers	Length (bp)	Reference
<i>B. ostreae</i>	F: CATTAAATTGGTCGGGCCGC R: CTGATCGTCTTCGATCCCC	300 bp	Cochennec <i>et al.</i> (2002)
	F: CAATGGTGCGTTCAACGAT R: GGGTTCGCGGTTGAATTTA	352 bp	Engelsma <i>et al.</i> (2010)
<i>B. exitiosa</i>	F: CATTAAATTGGTCGGGCCGC R: CTGATCGTCTTCGATCCCC	304 bp	Cochennec <i>et al.</i> (2000)
	F: CCGCACACGTTCTTCACTCC R: CTCGCGAGTTTCGACAGACG	1,090 bp	Le Roux <i>et al.</i> (2001)
<i>M. refringens</i>	F: TTTTGYTWGAGWGTTGCGAGATG R: CGAGTTTGCGAGTACCTCKAGAG	509 bp	Audemard <i>et al.</i> (2004)
<i>P. marinus</i>	F: CCGCTTTGTTTGGATCCC R: ACATCAGGCCTTCTAATGATG	490 bp	Kang <i>et al.</i> (2017)

Results and Conclusion

- A total of 1,860 oysters examined in this study, *B. ostreae*, *B. exitiosa*, *M. refringens*, and *P. marinus* were not detected from any individual oyster.
- Perkinsus olseni* was detected from 5 individuals out of 1,860 oysters collected from intertidal areas, and quantification by q-PCR indicated that the infection intensities were below 100 cells.
- The PCR also indicated that none of the oysters collected from subtidal long-lines were positive for *P. olseni*.

		Bonamiosis		Marteiliosis	Perkinsosis	
		<i>B. ostreae</i>	<i>B. exitiosa</i>	<i>M. refringens</i>	<i>P. marinus</i>	<i>P. olseni</i>
Commercial culture area	Spring	(-) 0/630	(-) 0/630	(-) 0/630	(-) 0/630	(-) 0/630
	Fall	(-) 0/630	(-) 0/630	(-) 0/630	(-) 0/630	(-) 0/630
Intertidal area	Spring	(-) 0/300	(-) 0/300	(-) 0/300	(-) 0/300	(-) 0/300
	Fall	(-) 0/300	(-) 0/300	(-) 0/300	(-) 0/300	(+) 5/300

- It is yet to be clear to define the detection of *P. olseni* in the oyster as infection.
- Further study is needed to confirm the *P. olseni* infection using histology.

Presenter: Hye-Mi Lee (hmlee@jejunu.ac.kr)
 Corresponding: Kwang-Sik Choi (skchoi@jejunu.ac.kr)